

Hydric and Edaphic Influence on Floristic Composition in an Altered Riparian Area

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ABSTRACT

Alteration of a natural ecosystem can physically, chemically and biologically affect a determined area. Therefore, this study aimed to evaluate the relation between the hydrology, soils and vegetation existing in an area changed by the implantation of a Hydroelectric Power Plant (HPP) in the municipality of Roque Gonzales, Rio Grande do Sul state. The study area was composed of three samples located downstream of the dam. This analysis was performed by the points method to verify the existence of similarity patterns between the vegetation, inundation levels and edaphic soil characteristics of each sample. As a result, greater floristic similarity was observed between the species as a relation of the spatial location with inundation levels. Thus, we can conclude that the vegetal establishment in the study area is under the direct influence of waterflow variation levels from the HPP, which contributes to defining species adaptability to these specific conditions, among others.

Keywords: environmental impacts, hydroelectric power plant, altered flow, riparian vegetation.

1. INTRODUCTION

Alteration in a hydrological regime caused by the installation of a dam generates (among other problems) social impacts, flooding of areas, formation of a reduced flow section (RFS) downstream of the work and impacts existing vegetation. Due to the fact that these areas present a lower flow rate than what is observed prior to establishing the work and the consequent reduction of existing flood spikes prior to implementing the project, a portion of distinct vegetation is formed with differentiated variability and proportion from the preexisting normal vegetation (New & Xie, 2008; Dwivedi, 2016; Bejarano et al., 2018), as well as differentiated soil characteristics (Stromberg, 2001; Silveira, 2006; Pereira et al., 2012).

Additionally (as discussed by Quadt et al., 2016), these altered areas can facilitate the entry of invasive species and compromise the diversity and integrity of the system, and should therefore be monitored and evaluated over time. In addition to these effects, fragmentation and terrestrialization tendencies can occur, as presented by Fanelli et al. (2015), as well as erosion and sedimentation in the area (New & Xie, 2008).

In this sense, the physical conditions of the environment play a key role in structuring communities in lotic environments. This is confirmed by Schneck & Hepp (2010) and Staes et al. (2010) who showed that the particular combination of several environmental factors such as current velocity, depth, bed substrate type, available light, organic matter and nutrients can determine the community that is associated with each habitat in a given watercourse.

Due to the fact that ciliary formations are connected to both the terrestrial and aquatic communities in an environment, they are influenced by both ecosystems. Thus, different types of soil are found in these areas, varying according to a greater or lesser degree of hydromorphism and the nature of the original material (Dalanesi et al., 2004; Curcio et al., 2007; Jacomine, 2009), as well as vegetation that is adapted to these conditions.

According to studies by Gonçalves et al. (2011), variations in fertility, soil acidity and altimetric elevation along a topographic gradient influence tree vegetation distribution; therefore, these characteristics directly influence the formation of local vegetation, its size,

structure and floristic composition. Thus, the study of variations that occur in these particular environments is essential, since they have a direct relation with the local flora and fauna, influencing and reflecting any changes that may occur in these areas (Ivanauskas et al., 1997; Lima et al., 2003; Kotchetkoff-Henriques et al., 2005; Carvalho et al., 2009).

In this context, this study is justified by the need of determining and evaluating the adaptation of the riparian vegetation to the altered environments, as well as the influence that these anthropic alterations cause in the environment, more specifically on the established and directly influenced vegetation. Therefore, the main objective of this study was to understand the variation and distribution of vegetation along a stretch of area altered by the implantation of a Hydroelectric Plant and its relation with inundation levels and soil edaphic characteristics. Hence, this study searched patterns in the vegetation establishment according to the existing edaphic variation for each inundation level, as well as possible similarities between the areas; thus confirming the influence of flow peaks (moisture) on forming and defining the vegetation established therein.

The importance of this evaluation is directly related to minimizing the environmental impacts generated by implementing an enterprise. It also supports the definition of possible measures to accelerate the restoration process of altered ciliary environments through selecting better-adapted species to the conditions observed there.

2. MATERIALS AND METHODS

The study area is located at the Passo do São João HPP, located on the Ijuí river, more specifically in the municipality of Roque Gonzales, Rio Grande do Sul state, 28°07'51" latitude to the south and 55°01'33" longitude, being part of the Ijuí river in the Northwest region of the state. The reservoir also covers the municipalities of São Luiz Gonzaga, São Pedro do Butiá and Rolador (Silveira, 2006), approximately 600 km from Porto Alegre (RS).

According to Streck et al. (2008), the region's soil is mainly classified in areas of agriculture, fields and forests. According to the Köppen classification system, the region presents a hot and humid temperate climate (Cfa) in all seasons, presenting hot summers. Regarding

precipitation in the region, data from 1988 to 2002 showed an annual average of 2210 mm, distributed throughout the year and with greater intensity in the months of March and April, September and October (Unijuí, 2003).

The Passo São João HPP is part of a cascade system in the Ijuí river, preceded by the São José HPP in the municipality of Cerro Largo. It is a waterborne power plant, meaning that its reservoir only has the function of maintaining the necessary slope for generating energy (Silveira, 2006).

With its implementation in 2010, a RFS of approximately 4 km in length was formed between the dam and the powerhouse with lower flow regime, where its flow regime flows in accordance with the requirements established in the licensing process of the enterprise. In this river section with altered flow, three sample areas (S1, S2, S3) were inserted to evaluate the vegetative and edaphic dynamics at different locations and flow characteristics (Figure 1).

Prior to the implantation of the hydroelectric plant, a varied range of ciliary vegetation along the canal formed this area, with agricultural influence

and a gradual slope embankment. The exception was sample area 1, which presented an abrupt slope in its altimetric levels with greater flow influence by the river due to the direct influence of the HPP, with the flow speed and the embankment eroded by water action.

The process of locating the sample areas, defining the flood levels and consequently the vegetation analysis lines on the slope were based on the flow data provided by Eletrosul from September 2011 to December 2013. The channel bathymetry through the ADCP (Acoustic Doppler Current Profiler) was used to calculate the flow in real time with a consequent adjustment using the daily flow data from 1958 to 2008 through the 75320000 fluviometric station, which is to say in the pre-establishment period of the dam.

This information was correlated through an evaluation of the key curve for the daily pre-dam flow data, adding the post-dam flow information and actual flows in the sample areas, thus defining the flood levels for each sample. This adjustment was made possible by the formula in Equation 1 (adapted from Silveira et al. 2010), as follows:

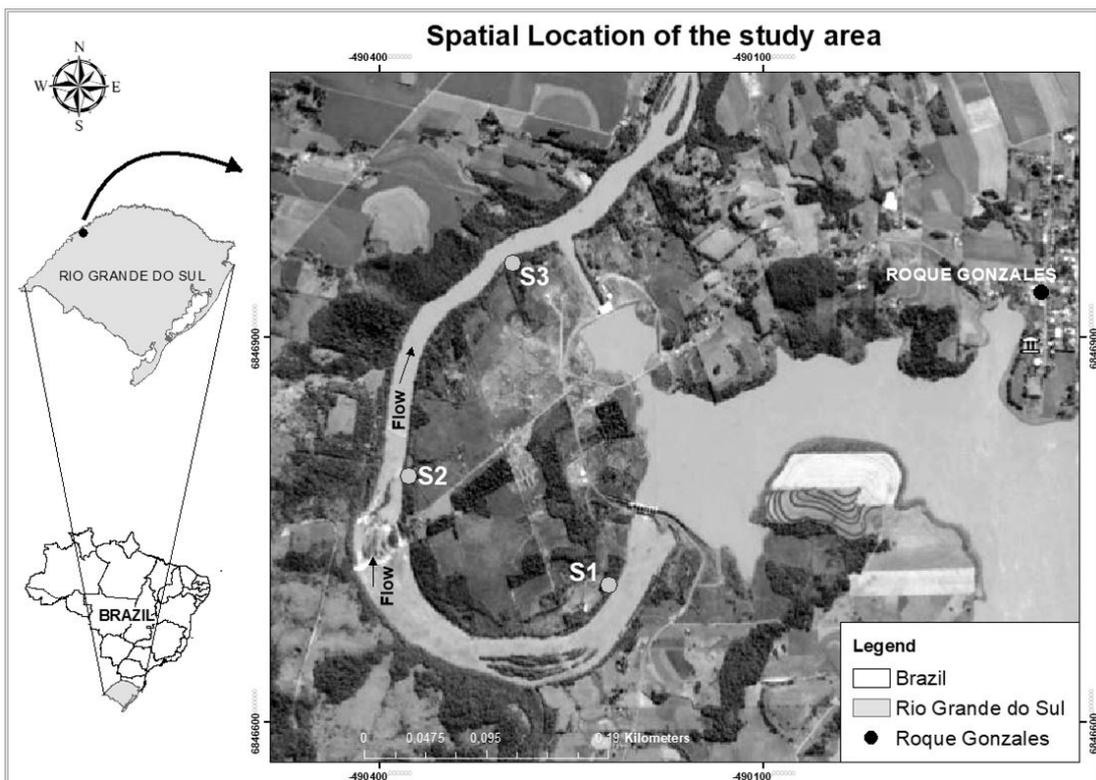


Figure 1. Spatial location of the study area and respective sample areas.

$$C_{est} = \left[\left(\frac{Q}{0.095} \right)^{1.534} + 57.62064 \right] \quad (1)$$

Where: C_{est} = estimated value of the dimension (cm);
 Q = Flow (m^3/s).

After the adjustments, the maximum levels of the flows in the RFS and their respective projected altimetric heights on the slopes of each area were estimated. The locating process of these sample units (SUs) involved systematic sampling by level intervals, which were demarcated in the field with the aid of a metric scale, a hose level and navigation GPS, with the altitudes of each SU then defined according to the estimated flow and flood characteristics.

2.1 Floristic survey

The points method adapted from Vieira & Pessoa (2001) was used for the vegetative analysis. Five SUs in each sample area were divided into 50-by-50 cm linear dimensions from "A" to "E" along the level curve and a length of 4.5 m, forming a network of 50 sample points per sample (10 sampling points per SA) and arranged in grid format, as according to Frizzo & Porto (2004). It should be noted that this methodology was used in order to not cause major disturbances in the environment, as well as to identify the small variations existing at the SU level, with the last level (dimension 5) representing a control for each sample, meaning not flooded.

The floristic survey of the herbaceous, shrub and liana strata, herein called life form analysis, was carried out at the species level using specialized literature, herbarium consultations and expert consultation, when necessary. The species classification followed the nomenclature proposed by Sobral et al. (2006), being classified by scientific nomenclature and by family.

The floristic comparison between the forest sections was carried out by elaborating a presence-and-absence matrix of the species, from which a dendrogram of similarity was constructed between all the sample areas and respective SU's, with all the species found according to the implemented methodology. The Jaccard similarity index was used according to Brower & Zar (1984) for elaborating the dendrogram, along with the Unweighted Pair Group Method with Arithmetic Mean (UPGMA).

2.2 Edaphic analysis

The soil samples were randomly collected in each SU with the aid of an auger in the 0 to 20 cm layer, collecting three single samples per level, forming one composed sample per level for each sample area. The samples were submitted to chemical and physical analysis in the laboratory of the Soils Department of the Rural Sciences Center – UFSM, with several parameters being analyzed such as: clay content, pH- H_2O , SMP index, Al Sat., Bases Sat., Clay, P, K, OM, Al, Ca, Mg, P, Cu, Zn, B, S and CEC pH7.

Interpretations of the chemical analysis reports were performed based on the Fertilization and Liming Manual for the states of Rio Grande do Sul and Santa Catarina (Tedesco et al., 2004). Statistical analysis of the data was performed by software R (R Core Team, 2016) through the Vegan library (Oksanen et al., 2016).

The canonical correspondence analysis (CCA) was used for the analysis and relation to the environmental variables, herein defined as the chemical variations of the soil. This allowed an analysis of the direct order of gradients and explained the species distribution in relation to the environmental variables, with the significance level assessed by the Monte Carlo permutation test (Braak & Prentice, 1988).

3. RESULTS

According to the floristic survey carried out in the study area (as presented in Table 1), 30 species and 20 distinct botanical families were identified, and these were classified as a function of the altimetric/flood (SU) levels in the section slope and sample areas.

According to Table 1 and in view of the sample evaluation, all sample areas showed species presence of the Boraginaceae, Euphorbiaceae, Fabaceae, Myrtaceae and Urticaceae families. In addition, only one botanical family (Euphorbiaceae) was verified when the nearest water levels (W1 and level1) of all sample areas were evaluated in the most unstable and saturated conditions of the study area.

According to Table 2, which followed the classification proposed by Tedesco et al. (2004), it is observed that Cation Exchange Capacity (CEC-pH7) and the Cu and Zn micronutrients were maintained at high concentrations in all the SUs of each sample

Table 1. Floristic/Phytosociological survey performed in the study area.

SAMPLE	LEVEL	SPECIES	FAMILY
S1	W1	<i>Eugenia uniflora</i> L.	Myrtaceae
		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae
		<i>Terminalia australis</i> Cambess.	Combretaceae
	level1	<i>Peplonia axillaris</i> Fontella & Rapini	Apocynaceae
		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae
		<i>Eugenia uniflora</i> L.	Myrtaceae
		<i>Parapiptadenia rigida</i> (Benth.) Brenan	Fabaceae
		<i>Sebastiania schottiana</i> (Müll.Arg.) Müll.Arg.	Euphorbiaceae
		<i>Ruellia angustiflora</i> (Ness) Lindau ex Rambo	Acanthaceae
		<i>Terminalia australis</i> Cambess.	Combretaceae
		<i>Cordia salicifolia</i> Cham.	Boraginaceae
	level2	<i>Eugenia uniflora</i> L.	Myrtaceae
		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae
		<i>Adenocalymma dusenii</i> Kraenzl.	Bignoniaceae
		<i>Sebastiania schottiana</i> (Müll.Arg.) Müll.Arg.	Euphorbiaceae
		<i>Ruellia angustiflora</i> (Ness) Lindau ex Rambo	Acanthaceae
		<i>Terminalia australis</i> Cambess.	Combretaceae
	level3	<i>Cordia salicifolia</i> Cham.	Boraginaceae
		<i>Eugenia uniflora</i> L.	Myrtaceae
		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae
		<i>Parapiptadenia rigida</i> (Benth.) Brenan	Fabaceae
		<i>Terminalia australis</i> Cambess.	Combretaceae
		<i>Ruellia angustiflora</i> (Ness) Lindau ex Rambo	Acanthaceae
		<i>Peplonia axillaris</i> Fontella & Rapini	Apocynaceae
		<i>Ruellia angustiflora</i> (Ness) Lindau ex Rambo	Acanthaceae
		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae
		<i>Cordia salicifolia</i> Cham.	Boraginaceae
	level4	<i>Terminalia australis</i> Cambess.	Combretaceae
		<i>Bromus catharticus</i> Vahl.	Poaceae
		<i>Pilea microphylla</i> Linn.	Urticaceae
		<i>Eugenia uniflora</i> L.	Myrtaceae
		<i>Plagiothecium curvifolium</i> Schlieph. ex Limpr.	Plagiotheciaceae
<i>Terminalia australis</i> Cambess.		Combretaceae	
<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.		Euphorbiaceae	
<i>Pilea microphylla</i> Linn.		Urticaceae	
<i>Ruellia angustiflora</i> (Ness) Lindau ex Rambo		Acanthaceae	
<i>Eugenia uniflora</i> L.		Myrtaceae	
level5	<i>Sebastiania schottiana</i> (Müll.Arg.) Müll.Arg.	Euphorbiaceae	
	<i>Adenocalymma dusenii</i> Kraenzl.	Bignoniaceae	
	<i>Bromus catharticus</i> Vahl.	Poaceae	
	<i>Cordia trichotoma</i> (Vell.) Arrab. ex Steud.	Boraginaceae	
	<i>Sebastiania commersoniana</i> (Baill.) L.B. Sm. & Downs	Euphorbiaceae	
	<i>Trema micrantha</i> (L.) Blume	Cannabaceae	
	<i>Sebastiania schottiana</i> (Müll.Arg.) Müll.Arg.	Euphorbiaceae	
	<i>Solanum paranense</i> Dusén.	Solanaceae	
	<i>Cordia trichotoma</i> (Vell.) Arrab. ex Steud.	Boraginaceae	
	<i>Sebastiania commersoniana</i> (Baill.) L.B. Sm. & Downs	Euphorbiaceae	
	<i>Eugenia pyriformis</i> Cambess.	Myrtaceae	
	level1	<i>Brachiaria plantaginea</i> (Link) Hitchc.	Poaceae
<i>Chrysophyllum marginatum</i> (Hook. & Arn.) Radlk.		Sapotaceae	
<i>Inga vera</i> Willd.		Fabaceae	
<i>Solanum paranense</i> Dusén.		Solanaceae	
S2	W1	<i>Eugenia uniflora</i> L.	Myrtaceae
		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae
		<i>Terminalia australis</i> Cambess.	Combretaceae
		<i>Peplonia axillaris</i> Fontella & Rapini	Apocynaceae
		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae
	level1	<i>Eugenia uniflora</i> L.	Myrtaceae
		<i>Parapiptadenia rigida</i> (Benth.) Brenan	Fabaceae
		<i>Sebastiania schottiana</i> (Müll.Arg.) Müll.Arg.	Euphorbiaceae
		<i>Ruellia angustiflora</i> (Ness) Lindau ex Rambo	Acanthaceae
		<i>Terminalia australis</i> Cambess.	Combretaceae

W1 = Water level.

Table 1. Continued...

SAMPLE	LEVEL	SPECIES	FAMILY	
S2	level2	<i>Pleurozium schreberi</i> (Willd. ex Brid) Mitt.	Hylocomiaceae	
		<i>Ageratum conyzoides</i> (L.) L.	Asteraceae	
		<i>Coutarea hexandra</i> (Jacq.) K. Schum.	Rubiaceae	
		<i>Chrysophyllum marginatum</i> (Hook. & Arn.) Radlk.	Sapotaceae	
		<i>Inga vera</i> Willd.	Fabaceae	
		<i>Serjania laruotteana</i> Cambess.	Sapindaceae	
		<i>Mikania cordifolia</i> (L.f.) Willd.	Asteraceae	
	level3	<i>Pleurozium schreberi</i> (Willd. ex Brid) Mitt.	Hylocomiaceae	
		<i>Inga vera</i> Willd.	Fabaceae	
		<i>Mikania cordifolia</i> (L.f.) Willd.	Asteraceae	
		<i>Brachiaria plantaginea</i> (Link) Hitchc.	Poaceae	
		<i>Terminalia australis</i> Cambess.	Combretaceae	
		<i>Serjania laruotteana</i> Cambess.	Sapindaceae	
		<i>Coutarea hexandra</i> (Jacq.) K. Schum.	Rubiaceae	
	level4	<i>Plagiothecium curvifolium</i> Schlieph. ex Limpr.	Plagiotheciaceae	
		<i>Sebastiania commersoniana</i> (Baill.) L.B. Sm. & Downs	Euphorbiaceae	
		<i>Chrysophyllum marginatum</i> (Hook. & Arn.) Radlk.	Sapotaceae	
		<i>Cordia salicifolia</i> Cham.	Boraginaceae	
		<i>Pleurozium schreberi</i> (Willd. ex Brid) Mitt.	Hylocomiaceae	
		<i>Pilea microphylla</i> Linn.	Urticaceae	
		<i>Eugenia involucrata</i> DC.	Myrtaceae	
		<i>Plagiothecium curvifolium</i> Schlieph. ex Limpr.	Plagiotheciaceae	
		<i>Coutarea hexandra</i> (Jacq.) K. Schum.	Rubiaceae	
		<i>Cordia trichotoma</i> (Vell.) Arrab. ex Steud.	Boraginaceae	
	level5	<i>Serjania laruotteana</i> Cambess.	Sapindaceae	
		<i>Cordia salicifolia</i> Cham.	Boraginaceae	
		<i>Bromus catharticus</i> Vahl.	Poaceae	
		<i>Eugenia uniflora</i> L.	Myrtaceae	
		<i>Eugenia involucrata</i> DC.	Myrtaceae	
		<i>Plagiothecium curvifolium</i> Schlieph. ex Limpr.	Plagiotheciaceae	
		<i>Pilea microphylla</i> Linn.	Urticaceae	
		<i>Cordia trichotoma</i> (Vell.) Arrab. ex Steud.	Boraginaceae	
<i>Chrysophyllum marginatum</i> (Hook. & Arn.) Radlk.		Sapotaceae		
<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.		Euphorbiaceae		
W1	<i>Bromus catharticus</i> Vahl.	Poaceae		
	<i>Inga vera</i> Willd.	Fabaceae		
	<i>Amphilophium crucigerum</i> (L.) L.G.Lohmann.	Bignoniaceae		
	<i>Coutarea hexandra</i> (Jacq.) K. Schum.	Rubiaceae		
	<i>Cordia trichotoma</i> (Vell.) Arrab. ex Steud.	Boraginaceae		
	<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae		
	<i>Serjania laruotteana</i> Cambess.	Sapindaceae		
	S3	level1	<i>Amphilophium crucigerum</i> (L.) L.G.Lohmann.	Bignoniaceae
			<i>Coutarea hexandra</i> (Jacq.) K. Schum.	Rubiaceae
			<i>Cordia trichotoma</i> (Vell.) Arrab. ex Steud.	Boraginaceae
<i>Inga vera</i> Willd.			Fabaceae	
<i>Adenocalymma dusenii</i> Kraenzl.			Bignoniaceae	
level2		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae	
		<i>Coutarea hexandra</i> (Jacq.) K. Schum.	Rubiaceae	
		<i>Eugenia uniflora</i> L.	Myrtaceae	
		<i>Cordia trichotoma</i> (Vell.) Arrab. ex Steud.	Boraginaceae	
		<i>Inga vera</i> Willd.	Fabaceae	

W1 = Water level.

Table 1. Continued...

SAMPLE	LEVEL	SPECIES	FAMILY
S3	level3	<i>Amphilophium crucigerum</i> (L.) L.G.Lohmann.	Bignoniaceae
		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae
		<i>Coutarea hexandra</i> (Jacq.) K. Schum.	Rubiaceae
		<i>Eugenia involucrata</i> DC.	Myrtaceae
		<i>Pilea microphylla</i> Linn.	Urticaceae
	level4	<i>Coutarea hexandra</i> (Jacq.) K. Schum.	Rubiaceae
		<i>Amphilophium crucigerum</i> (L.) L.G.Lohmann.	Bignoniaceae
		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae
	level5	<i>Ageratum conyzoides</i> (L.) L.	Asteraceae
		<i>Marchantia paleacea</i> Bertol.	Marchantiaceae
		<i>Peplonia axillaris</i> (Vell.) Fontella & Rapini	Apocynaceae
		<i>Actinostemon concolor</i> (Spreng.) Müll.Arg.	Euphorbiaceae
		<i>Sorocea bonplandii</i> (Baill.) W.C. Burger, Lanjouw & Boer	Moraceae
		<i>Plinia rivularis</i> (Cambess.) Rotman	Myrtaceae

W1 = Water level.

Table 2. Chemical characteristics of the soil samples (layer 0-20 cm) collected in 3 reduced flow section (RFS) SUs of the HPP, Passo São João, Rio Grande do Sul.

Sample Level	pH _{H₂O}	Ca	Mg	Al	CECef	AlSat.	BasesSat.	SMP	OM	Clay	S	K	CEC-pH7	Cu	Zn	B
		---cmol /dm ³ ---			---(%)---		--(%)--		mg/dm ³	cmol /dm ³						
S1L1	5.4	14.9	8.5	0.2	23.9	0.8	87	6.2	0.7	32	13	92	27.2	14.8	4.4	0.2
S1L2	5.4	15.9	9.1	0.1	25.3	0.4	86.5	6.1	1	30	8.3	64	29.1	13.1	4.6	0.1
S1L3	5.4	13.6	7.8	0.1	21.7	0.5	84.5	6.1	1.2	31	7	68	25.5	13.4	3.8	0.2
S1L4	5	11.7	7.2	0.1	19.2	0.5	75.4	5.7	2.1	30	13	72	25.3	14.4	6.4	0.2
S1L5	4.9	11.5	7.4	0.3	19.3	1.6	71.2	5.5	1.9	31	13	72	26.7	13.3	5.4	0.2
S2L1	5.5	11.9	7.4	0	19.4	0	88.7	6.5	0.9	24	8	48	21.9	11.5	5.3	0.2
S2L2	5.4	9.8	5.9	0.1	15.9	0.6	81.6	6.2	1	21	8.1	32	19.3	11.5	5.1	0.1
S2L3	5.5	10.8	7.1	0	18.1	0	82.2	6.1	1.6	25	14	60	22	12	4.6	0.1
S2L4	5.1	9.5	6.7	0.3	16.7	1.8	77	5.9	1.9	25	17	68	21.3	11	3.9	0.4
S2L5	4.7	6.7	6	1	13.9	7.2	56.9	5.3	2.3	24	17	64	22.6	11.2	3.7	0.1
S3L1	5.5	15.6	8.7	0	24.5	0	84.6	6	2.2	31	16	40	28.9	18	8.6	0.1
S3L2	5.5	15.6	8.4	0	24.2	0	84.5	6	2.2	31	11	44	28.6	17.9	8.8	0.1
S3L3	5.5	15.5	6.8	0	22.4	0	80.2	5.8	2.4	34	15	28	27.9	18.5	8.7	0.1
S3L4	5.6	16.9	8.1	0	25.1	0	86.6	6.1	2.2	31	15	40	29	17.6	9	0.1
S3L5	5.7	18.3	9.2	0	27.6	0	87.7	6.1	2.6	30	16	44	31.5	16.6	9.6	0.1

area (mean of 21.14 cmolc/dm³, 14.32 mg/dm³ and 6.12 mg/dm³, respectively), with B concentration found being classified as average in most levels (average of 0.11 mg/dm³), with the exception of level 4 in sample area 2 (S2), which presented high concentrations of this element (0.4 mg/dm³). For the analyzed macronutrients (P, K, Ca, Mg and S), they presented mean concentrations of: 2.8 mg/dm³, 55.7 mg/dm³, 13.21 cmolc/dm³, 7.62 cmolc/dm³, and 12.76 mg/dm³,

thus being classified as very low and low for P and K respectively, and high for the other elements.

Regarding organic matter (OM), it presented a low concentration in the soil (mean of 1.7 m/v%), which demonstrates the low availability of this element to the plants in the considered area. For pH in water, an average of 5.34 was found in its index, which ranks it as low pH or acid. Furthermore, an average of 80.9% was verified regarding the percentage of base saturation in

the analyzed samples, which classifies it as high on the base saturation index. However, this percentage drops to 0.89% for aluminum saturation, thus representing a very low index for this soil. The dendrogram shown in Figure 2 shows the floristic relationships generated by the grouping analysis of the species found in each sample area and SU by means of the group average (UPGMA), with the similarity then being greater than 50%, as highlighted.

For the four highest similarity indexes (defined by the highlighted horizontal line), it was observed that these clusters were separated according to the location of the sample area and with the tendency to be in the nearest SUs. The subgroups presenting the greatest similarity among each other were: S1_L1 and S1_L3, S1_L4 and S1_L5, located in sample area 1; and S3_L1 and S3_WL, S3_L3 and S3_L4, located in sample area 3. This means that there was a high similarity between the plant species found at these levels and sites.

In these groupings, there was a greater affinity between the establishment/presence of certain species such as *Parapiptadenia rigida* and *Peplonia axillaris*; *Marchantia paleacea* and *Plinia rivularis*; *Sebastiania commersoniana* and *Solanum paranense*; *Eugenia involucrata* and *Pilea microphylla*; and finally *Coutarea hexandra* and *Inga vera*.

The co-optic correlation coefficient (ccc) proposed by Sokal & Rohlf (1962) presented a value of 0.88, which indicates suitability in the grouping method used (Rohlf, 1970).

The ordering diagram of the plots in the first axis was generated by means of Canonical Correspondence Analysis (CCA) (Figure 3). CCA shows the division into groups with positive and negative eigenvalues, allied to the axes being sample areas (S1, S2 and S3) and their respective SUs with the floristic information and soil chemical characteristics (environmental variables) for each site. Eigenvalues of 0.84 and 0.72 were found for the CCA1 and CCA2 axes, respectively, which represented a percentage explanation of 20.9% and 18% of the data variance for these axes and correlated variables.

With this analysis, one can observe a clear division of the sample area 1 (S1) in relation to the others. This area showed a strong correlation between the presence of certain plant species such as *Parapiptadenia rigida*, *Sebastiania schottiana*, *Ruellia angustiflora* and *Terminalia australis*, among others, with soils containing higher concentrations of K, Al, Al Sat. and clay content.

In addition, species such as *Brachiaria plantaginea*, *Eugenia pyriformis*, *Sebastiania commersoniana* and *Solanum paranense* presented a higher correlation with the environmental variables: Base Sat., SMP and

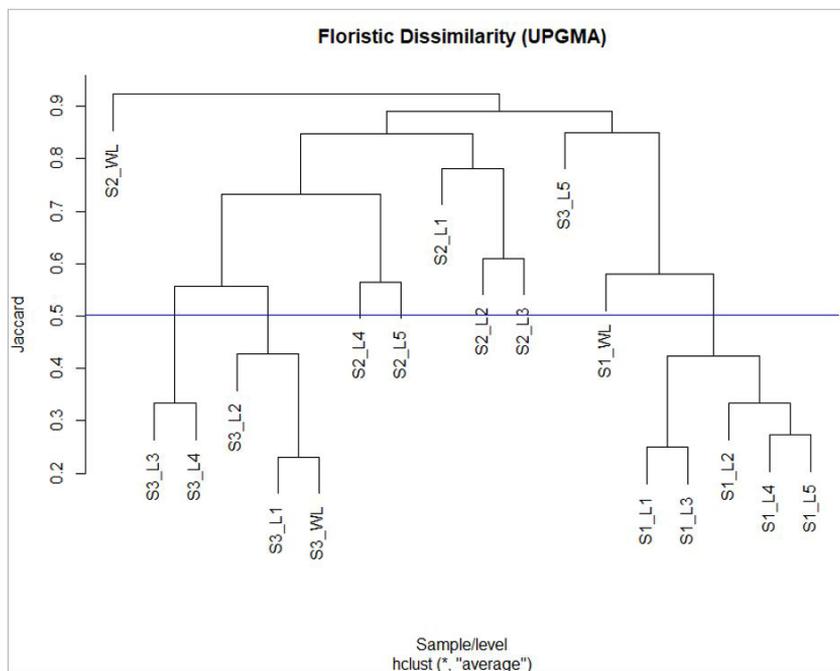


Figure 2. Dendrogram obtained through Jaccard index (UPGMA), based on the vegetal species found in study area.

In this sense, it is possible to observe (via CCA) the ordering and grouping of plant species as a function of flood levels, spatial location along the RFS and soil chemical characteristics, which demonstrated the direct connection and complexity of these variables. In turn, such variables are responsible for the adaptation, establishment and permanence of these species under specifically induced or naturally occurring conditions, together with other factors and elements that act directly and indirectly in the environment.

Regarding the behavior of soil chemical elements in the analyzed areas and their availability to plants, this study confirms the results observed by Rodrigues et al. (2007). The authors show that an increase of soil acidity is inversely correlated with the levels of Al, K, Ca and Mg. Thus, it can be verified that the establishment of some plant species such as *Brachiaria plantaginea*, *Sebastiania commersoniana*, *Solanum paranense*, among others, in environments with higher pH, meaning with less availability of certain plant elements (Marschner, 1995), demonstrates the low nutritional requirement of these plants and their high adaptability under “restrictive” conditions imposed by this environment.

5. CONCLUSIONS

In this study, there was a tendency of greater similarity between plant species inserted in similar levels to each other, meaning with a direct influence of the RFS flow. The influence of the flow rate and soil characteristics have a direct influence on the existence and establishment of plant species according to their adaptation to the imposed conditions, even in small distances between them. In addition, a tendency in grouping by sample area was generally observed. The hypothesis for this behavior (variation along the stretch) is that the influence of specific conditions/factors in each area (meaning the flow rate characteristics in each section, terrain/slope, and soil chemical conditions, among others) acts directly in the establishment of plant species.

Moreover, the presence of certain botanical families in specific areas and levels was verified. For example in the case of the S3 sample area and level5 (S3L5), which was the only one that presented species of the Marchantiaceae and Moraceae families. Therefore there is a tendency of floristic differentiation in

higher levels (with less influence of floods) that can be explained by possible less adaptability to the edaphic and humidity conditions of the environments closer to the river channel.

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